



INVESTIGATION FOR IN VITRO ANTIBACTERIAL ACTIVITY OF *CITRUS SINENSIS* L. AGAINST *PROPIONIBACTERIUM ACNES*

Manisha Pandey., Afifa Qidwai., Anand Pandey., Shashi Kant Shukla., Rajesh Kumar., Ashutosh Pathak and Anupam Dikshit*

Biological Product Laboratory, Department of Botany, University of Allahabad, Allahabad-211002, India

ARTICLE INFO

Article History:

Received 20th May, 2017

Received in revised form 12th

June, 2017 Accepted 6th July, 2017

Published online 28th August, 2017

Key words:

Acne vulgaris; *Propionibacterium acnes*; *Citrus sinensis* L.; MIC; IC₅₀; etc.

ABSTRACT

Aim: The present study intends to observe and compare the bactericidal activity of the crude extract of *C. sinensis* L. peels in different solvent and essential oil against *P. acnes* and their phytochemical analysis.

Background of the Study: Acne is a chronic, inflammatory disease, of pilosebaceous unit. Hormonal imbalance, bacterial infection, stress, improper feeding habits and cosmetic applications are major factors considered to cause the development of acne. *P. acnes*, a gram-positive, anaerobic bacterium, has been recognized as the key factor for development of acne. Traditionally, antibiotics and hormonal doses have been applied to treat acne. However, these treatments come along with severe side effects and drug resistance.

Methodology: The phytochemical analysis of alkaloids test, flavonoids test, steroids test and saponins test were analyzed. The antibacterial activity of these extracts and essential oil were screened in accordance to globally accepted CLSI guidelines. Further, herbal extracts and oil activities were also compared with synthetic Tetracycline which was used as a standard.

Results: The results (mg/ml) indicate that ethanolic extracts of *C. sinensis* L. (MIC: 0.074, IC₅₀: 0.155) as most promising form of crude extract and higher antibacterial properties as compared to petroleum ether extracts (MIC: 0.722, IC₅₀: 0.658), peel essential oil (MIC: 1.357, IC₅₀: 1.301) and acetone extracts (MIC: 1.390, IC₅₀: 1.328), respectively. It therefore, provides a promising herbal supplement for the management of Acne vulgaris and that to from a waste resource.

Copyright©2017 Anupam Dikshit et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Acne is a chronic, inflammatory disease, clinical manifestation involves from a mild comedonal form to severely visible cystic acne on face, chest, and back. Hormonal imbalance, bacterial infection, stress, improper feeding habits, and cosmetic applications are major factors considered to cause the development of acne [1]. *Propionibacterium acnes*, a gram-positive, anaerobic bacterium, has been recognized as the key factor for development of acne [2]. Traditionally, antibiotics and hormonal doses have been applied to treat acne [3,4]. However, these treatments come along with severe side effects and drug resistance [5,6]. Antibiotics which have been known to suppress *P. acnes* are becoming less effective probably because of the emergence of antibiotic-resistant strains [7,8,9,10]. Quest for an effective treatment that is well received by patients is still a challenge. Phytotherapeutic approaches for treatment with highly antibacterial herbal

supplements/ substitutes and minimum side effects have recently become the thrust for research in this field.

Essential oils and herbal extracts have been widely used in remediation of several diseases from decades [11, 12]. Their anti-microbial and anti-oxidant properties open the hidden possibilities that these natural resources offer [13, 14] such as use of plant extracts and oil for cure of acne [15]. Several plants are being utilized since prehistoric times for the same purpose; *C. sinensis* L. is one of them.

C. sinensis L. (Orange, Family- Rutaceae) is one of the most important commercial fruit crops grown in nearly all countries of the world.^[16] It is rich in nutrients as well as in phytochemicals such as flavanones, polyphenols, anthocyanins and hydroxycinnamic acids which are beneficial in the remedy of high cholesterol, irritation, itching, inflammation and complications related to Acne vulgaris. Among the nutrients found in orange peel essential oil, the most important are limonoid compound, which can be effectively used as an alternative of synthetic drug.^[17] Limonoids exhibit scavenging activity for superoxide anion

*Corresponding author: Anupam Dikshit

Biological Product Laboratory, Department of Botany,
University of Allahabad, Allahabad-211002, India

radicals and ultimately inhibits *P. acnes* induced secretion of oil. Moreover, there is a lack of information regarding the inhibitory effects of orange peel extracts with reference to different solvents and their oil.

The present work focuses on the comparative study of antibacterial effects of *C. sinensis* L. peels oil and extracts in various solvents viz; ethanol, acetone and petroleum ether against *P. acnes* by Broth Micro dilution method recommended by Clinical and Laboratory Standards Institute (CLSI).

METHODOLOGY

Collection of bacterial Culture - *P. acnes* culture in lyophilized form was procured from Microbial Type Culture Collection (MTCC-1951), Chandigarh, India. It was revived and maintained on Anaerobic Blood Agar medium supplemented with sheep blood. Anaerobic environment was provided by Anaxomate Advance Instrument (Mart Microbiology B.V.) and kept in incubation for 48 hours at $35 \pm 2^\circ\text{C}$. (Figure 1).

Essential oil extraction-The peels of *C. sinensis* L. were collected from the juice shops and washed thoroughly with tap water and dried in shade to maintain their active constituents. Consequently (200g) peels were chopped and loaded in the Clevenger apparatus for oil extraction through hydro-distillation.^[18] The extracted oil (Figure 1) was stored at 4°C for subsequent studies.



Figure 1 A-Culture of *P. acnes* on Anaerobic blood Agar media B-
C. sinensis L. peel essential oil

Preparation of extracts- After drying, 10 grams of *C. sinensis* L. peel was weighed chopped and soaked overnight in 100 ml of 50% ethanol, petroleum ether and acetone respectively. All organic solvents and other chemicals were of analytical grade. Subsequently, the extract solutions were filtered by Whatman filter paper no.1, and followed by evaporation under vacuum and optimal temperature in rotatory evaporator apparatus to

obtained crude extracts. Further the extracts had been dried completely and stored at 4°C for further use. Percentage yield was calculated as below:

Percentage Yield = Amount of extract (in gm)/amount of sample (in gm) $\times 100$



Figure 2 Preparation of various solvent extracts through Rotatory evaporator

Phytochemical analysis

Alkaloids Test-2 ml *C. sinensis* L. peel filtrate were mixed with 1% HCl and about 6 drops of Mayor's reagents. A Creamish or pale yellow precipitate indicates the presence of alkaloids.^[19]

Flavonoids Test: 2 ml filtrate was added to conc. HCl and magnesium ribbon. Pink-tomato red colour indicates the presence of Flavonoids [19].

Steroids Test: 2 ml of acetic anhydride was added to 0.5 gram ethanolic extract of sample and added 2 ml H_2SO_4 . The color changed from violet to blue or green in sample indicate the presence of steroids [19].

Saponins Test: For detection of saponins Froth test is being used. 1 g of the sample weighed and put into a conical flask in which 10 ml of sterile distilled water was added and boiled for 5 min. The mixture was filtered and 2.5 ml of the filtrate was added to 10 ml of sterile distilled water in a test tube. The test tube was sealed and shaken vigorously for about 30 sec. It was then allowed to stand for half an hour. Honeycomb froth (foam) indicated the presence of saponins [19].

Antibacterial assay

The susceptibility of the *P. acnes* against *C. sinensis* L. peel crude extracts was assayed using CLSI recommended broth microdilution method[20] Freshly prepared Muller Hinton Broth medium was used for the assay. A 50 mg/ml stock solution of extracts and synthetic drug were prepared in 950 mg/ml dimethyl sulfoxide (DMSO) and homogenized by using vortex for 4-5 min. Bacterial inoculum was prepared as per 0.5 McFarland standards [21]. Tetracycline, as a standard drug, was used for antibacterial assay. The experiment was performed according to CLSI guidelines in flat bottom sterile 96-well micro-titre plates and cultured overnight. Initial dispensing of 100 μl medium (MHB) was followed by the addition of 90 μl and 80 μl of MHB in columns 3 and 4 respectively. Further, 10 μl and 20 μl of drugs (orange peel extracts and oil) were added in each well of columns 3 (sample control) and 4 (dilution well) respectively. Following this, a two-fold serial dilution was done from 4th column wells (2.5mg/ml) to 11th column wells (0.02mg/ml), to which 100 μl initially prepared bacterial inoculums was added to make up 200 μl .

Column 1 contained media and formaldehyde to serve as negative control. Column 12 was taken as the positive control (O. D. Control), which contains 100 μl medium and 100 μl inoculum (Fig: 4). The experimental 96 well plates were kept in the anaerobic jar of Anaxomate. The jar was then incubated

Investigation for in vitro antibacterial activity of citrus sinensis l. Against propionibacterium acnes

in CO₂ incubator (Galaxy 170 S New Brunswick, USA) for 24-48 hr.

The results of our study clearly portray significant antibacterial with reference to the MICs as well as IC₅₀

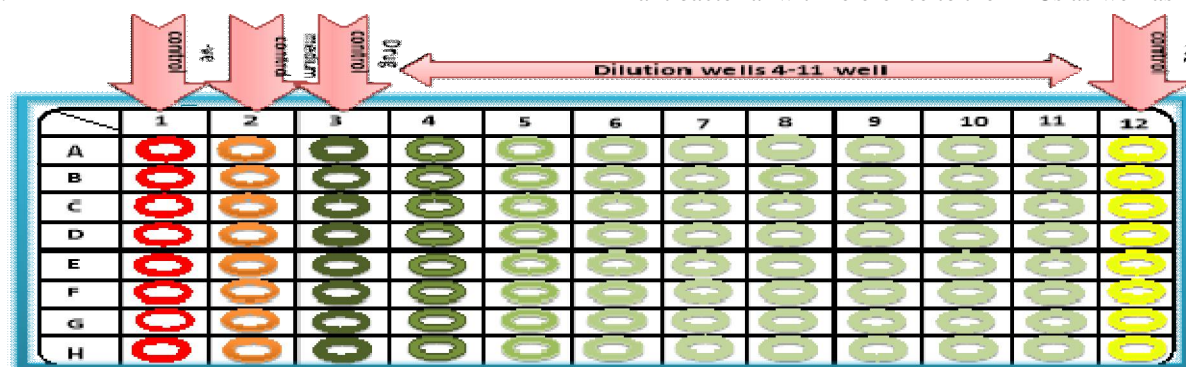


Figure 3 A detailed diagram of used 96 well plates in CLSI recommended broth microdilution technique

The turbid metric method of MIC and IC₅₀ calculation was used by Spectramax Plus 384 (Molecular Devices, USA) spectrophotometer at 492 nm. The tests were implemented in duplicates. For herbal extracts and oil, comparative inhibition percent of bacterium inoculum in media was calculated by using the following formula.

$$\% \text{ Inhibition} = \frac{(\text{O.D. Control} - \text{O.D. treatment})}{\text{O.D. control}} \times 100 \text{ [22].}$$

RESULTS

In our current study the preliminary quantitative analyses of the *C. sinensis* L. peels extracts (ethanol, acetone and petroleum ether) and oil were executed to analyze antibacterial and phytochemical properties against *P. acnes*. Presence of phytochemicals in *C. sinensis* L. was pinpointed after screening peel crude extracts filtrate through ethanol, petroleum ether and acetone (Table1). It has been found that flavanones and many polymethoxylated flavones were present in the peels [23,24]. Important flavonoids found in *C. sinensis* L. are limonene, hesperidin, narirutin, naringin and eriocitrin [25].

Table 1 Phytochemical analysis of *C. sinensis* L. peel extracts

Secondary metabolites	Reagent	Observation	Results
Alkaloids	Meyer	Whitemist formation	+
Flavonoids	Siandin test	Orange solution	+
Steroids	Acetic anhydride+ H ₂ SO ₄	Blue Solution	+
Saponin	H ₂ O	Honeycomb froth	+

The yields of herbal extracts and oil reveal the difference in the findings of the present observations. Maximum yield occurred in oil; percent yield of ethanol extract was 4 times more than the Petroleum ether solvent extract and percent yield of acetone is very less. This variation in the yield of the aforesaid solvent extracts is due to disparity in solubility of different plant compound with solvents.

Table 2 Antibacterial activity of *C. sinensis* L. peel ethanolic, petroleum ether, acetone extracts, Oil and Tetracycline against *P. acnes* (mg/ml).

Pathogenic microbe	Bactericidal activity (IC ₅₀ and MIC)									
	<i>Citrus sinensis</i> L.								Standard drug (Tetracycline)	
	Ethanolic extract		P.Ether extract		Acetone extract		Essential oil		IC ₅₀	MIC
<i>P. acnes</i>	IC ₅₀	MIC	IC ₅₀	MIC	IC ₅₀	MIC	IC ₅₀	MIC	0.013	0.028
	0.155	1.357	0.658	0.658	1.328	0.074	1.301	1.357		

(mg/ml) values through 96 well microtitre plate (CLSI recommended broth micro dilution method. Antibacterial effects of crude ethanolic, acetone, petroleum ether extracts and essential oil of the peels of *C. sinensis* L. was evaluated against *P. acnes*. These drugs clearly reflected the significant activity against the bacterium. Among these drugs ethanolic extracts of *C. sinensis* L. demonstrated higher activity with (MIC: 0.074, IC₅₀:0.155) compared to petroleum ether (MIC: 0.722, IC₅₀:0.658) and oil (MIC: 1.357, IC₅₀:1.301). However, acetone extracts showed least efficacy. All these results were compared with Tetracycline (MIC: 0.028, IC₅₀: 0.013) (standard antibiotic). The results showed that tetracycline had stronger activity than tested extracts as shown in Table 2.

DISCUSSION

Above results clearly indicates the high bactericidal activity of ethanolic extract against the tested anaerobic bacteria is attributed to the presence of high alkaloids, saponins, tannins, steroids, phenols and Flavonoids. However, difference in the activity of different extracts may be because of the difference in solubility of phytochemicals in different solvents. Ethanol maximizes the bioavailability of phytochemicals such as tannins [26], polyphenol, polyacetylenes [27, 28], flavonol [29], terpenoids [30], sterol [31], alkoloides [32], which is not in case with petroleum ether which extracted flavonoids [34] and polyphenols where as acetone extracted only flavonols. These classes of secondary metabolites are known to possess antibacterial property. It has been reported that an important aspects of this class of compounds is their pharmacological activity as scavengers for superoxide anion radicals like O²⁻, OH., peroxy radicals and singlet oxygen[35]. This property of flavonoids protects intracellular and extracellular structures of skin [36,37] which is highly infected by *P. acnes*. MIC and IC₅₀ value of ethanolic extracts of *C. sinensis* L. peels obtained through CLSI method is very low as compared to petroleum ether, oil and acetone which is also clearly shows that ethanolic extracts of *C. sinensis* L. is highly active against the treatment of acne vulgaris with less side effects.

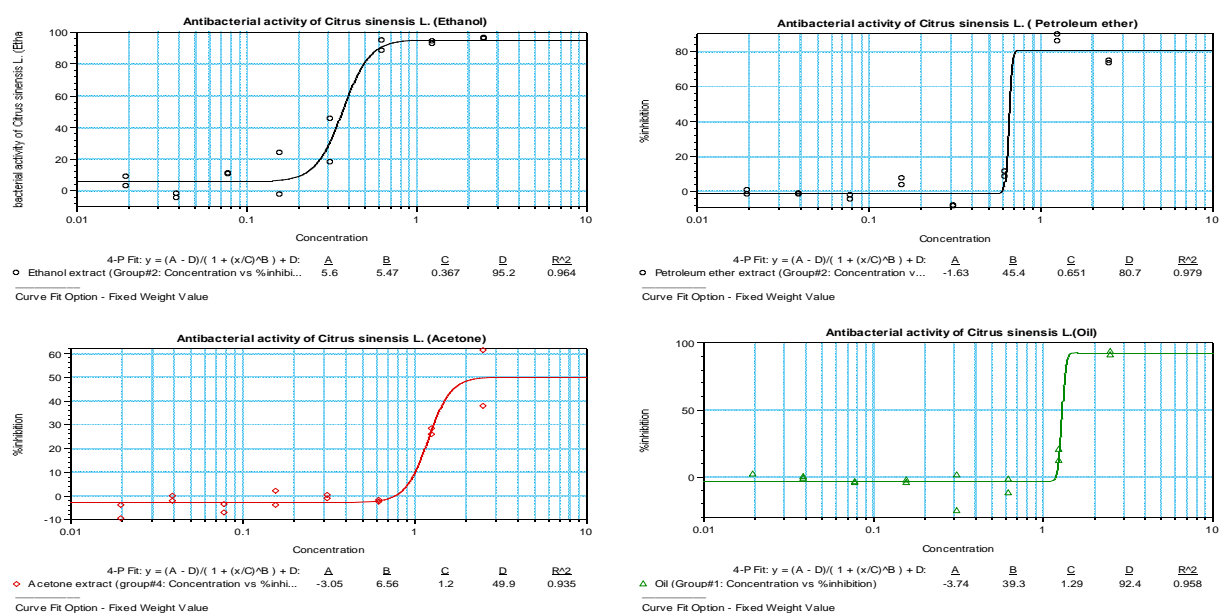


Figure 4 Graph showing antibacterial activity of *C. sinensis* L. peel against *P. acnes*, at 24 hours. (A) Ethanolic extract; (B) Petroleum ether extract; (C) Acetone extract; and (D) Essential oil

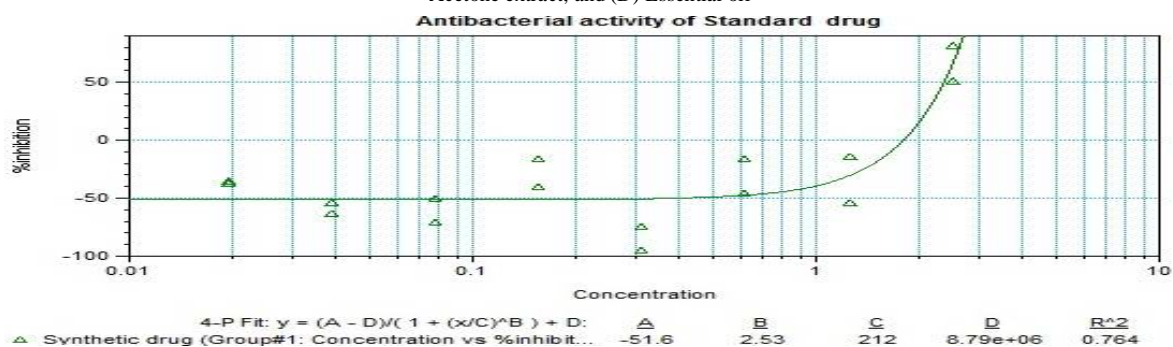


Figure 5 Graph showing antibacterial activity of standard drug (Tetracycline) against *P. acnes*, at 24 hours.

CONCLUSIONS

Therapeutic approaches based on the herbal medication is considered safer than synthetic medicines because their precursors are associated with side effects such as contact allergy, local irritation, itching, redness, skin peeling, etc. Present finding has evaluated the antibacterial activity against the *P. acnes* and phytochemical constituents in *Citrus* peels extracts obtained from different solvents. Ethanolic extracts of peels was found to exhibit higher antibacterial activity as compared to extracts from petroleum ether, acetone and essential oil. This is not all; further research should focus on isolation of novel active compounds from the peels extract and essential oil and to assess their bioactivities, as it is necessary to introduce naturally safe phytochemical compounds that can suppress the microbial growth. Further, there is a need of *in vivo* studies to determine the acceptability and safety of these drugs. Further, evaluation performed with pure compounds is required for definite inference of the bioactive compounds contributing to the antimicrobial activity.

Acknowledgements

Thanks are due to the Head, Department of Botany, University of Allahabad, Mr. Rick Z for anaerobic jar, Motilal Nehru Medical College, for anaerobic culture; and UGC for financial support.

References

1. Park J, Lee J, Jung E, Park Y, Kim K, Park B, Jung K, Park E, Kim J, Park D. In vitro antibacterial and anti-inflammatory effects of honokiol and magnolol against *Propionibacterium* sp. *Eur. J. Pharmacol* 2004; 496:189-195.
2. Jappe U. Pathological mechanisms of acne with special emphasis on *Propionibacterium acnes* and related therapy. *Acta Derm. Venereol.* 2003; 83: 241-248.
3. Poulin Y. Practical approach to the hormonal treatment of acne. *J. Cutan. Med. Surg.* 2004; 4:6-21.
4. Tan A W, Tan H H. Acne vulgaris: a review of antibiotic therapy. *Expert Opin. Pharmacother* 2005; 6:409-418.
5. Leyden J. Antibiotic resistance in the topical treatment of acne vulgaris. *Cutis* 2004; 73:6-10.
6. Yemisci A, Gorgulu A, Piskin S, Effects and side-effects of spironolactone therapy in women with acne. *J. Eur. Acad. Dermatol. Venereol.* 2005; 19:163-166.
7. Leyden JJ. Antibiotic resistant acne. *Cutis* 1976; 17:593-6.
8. Leyden JJ, McGinley KJ, Cavalieri S, Webster GF, Mills OH, Kligman AM. *Propionibacterium acnes* resistance to antibiotics in acne patients. *J Am Acad Dermatol* 1983; 8:41-5.
9. Eady EA, Cove JH, Holland KT, Cunliffe WJ. Erythromycin resistant propionibacteria in antibiotic

- treated acne patients: Association with therapeutic failure. *Br J Dermatol* 1989; 121:51-7.
10. Eady EA, Jones CE, Tipper JL, Cove JH, Cunliffe WJ, Layton AM. Antibiotic resistant propionibacteria in acne: Need for policies to modify antibiotic usage. *BMJ* 1993; 306: 555-6.
 11. Buckle J. Use of aromatherapy as a complementary treatment for chronic pain. *Altern Ther Health Med* 1999; 5:42-51.
 12. Sylvestre M, Pichette A, Longtin A, Nagau F, Legault J. Essential oil analysis and anticancer activity of leaf essential oil of *Croton flavens* L. from Guadeloupe. *J. Ethnopharmacol* 2006; 103, 99-102.
 13. Mimica-Dukic N, Bozin B, Sokovic M, Simin N. Antimicrobial and antioxidant activities of *Melissa officinalis* L. (Lamiaceae) essential oil. *J. Agric. Food. Chem* 2004; 52, 2485-2489.
 14. Sylvestre M, Legault J, Dufour D, Pichette A. Chemical composition and anticancer activity of leaf essential oil of *Myrica gale* L. *Phytomedicine*. 2005; 12:299-304.
 15. Lertsatitthanakorn P, Taweechaisupapong S, Aromdee C, Khunkitti W. In vitro bioactivities of essential oils used for acne control. *Int. J. Astrobiol* 2006; 16:43-49.
 16. Sokovic M, Petar D, Marin, Dejan Brkic, Leo J L D, Griensven Van. Chemical Composition and Antibacterial Activity of Essential Oils of Ten Aromatic Plants against Human Pathogenic Bacteria Global Science Books Food 2007;1.
 17. Kim SS, Baik JS, Oh TH, Yoon WJ, Lee NH, Hyun CG. Department of Chemistry, Cheju National University PMID: 18838824 [PubMed-MEDLINE].
 18. Clevenger J F. Apparatus for the determination of volatile oil. *J Am Pharm. Assoc.*, 1928; 17: 3.
 19. Haroen U, Marlida Y, Mirzah, Budianyah A. Extraction and Isolation Phytochemical and Antimicrobial Activity of Limonoid Compounds from Orange Waste Juice. *Pakistan Journal of Nutrition* 2013; 12 (8): 730-735.46-346.
 20. Methods for antimicrobial susceptibility testing of Anaerobic bacteria (8th ed.), Approved standard, M7-A5. Pennsylvania: Wayne. NCCLS; 2012.
 21. McFarland J. The nephelometer, an instrument for estimating the number of bacteria in suspension used for calculating the opsonic index and for vaccines. *J. Am. Med. Assoc.* 1907; 49, 1176-1178.
 22. Pathak A, Shukla SK, Pandey A, Mishra RK, Kumar R, Dikshit A. In vitro antibacterial activity of Ethno medicinally used lichens against three wound infecting genera of Enterobacteriaceae. *Proc. Natl. Acad. Sci. India B Biol. Sci.*2015;DOI:10.1007/ s40011-015-0540-y.
 23. Tao NG, Hu ZY, Liu Q, Xu J, Cheng YJ, Guo LL, Guo WW, Deng XX. Expression of phytoene synthase gene is enhanced during fruit ripening of navel orange (*Citrus sinensis*) *Plant Cell Rep.* 2007; 26: 837- 843.
 24. M M Ahmad, Rehman ur Salim, Iqbal Z, Anjum FM, Sultan J I, Genetic variability to essential oil composition in four citrus fruit species. *Pak. J. Bot* 2006; 38(2): pp.319-324.
 25. Ashok kumar K., Narayani M., Subanthini A. and Jayakumar M. Antimicrobial Activity and Phytochemical Analysis of Citrus Fruit Peels - Utilization of Fruit Waste. *International Journal of Engineering Science and Technology (IJEST)* 2011; ISSN: 0975- 5462 Vol. 3 No.
 26. Francis, F.J Wiley encyclopedia of food science and technology Wiley Interscience, USA 2000; Vol. 4:pp. 2449-2467.
 27. Silva O, Duarte A, Pimental M, Viegas S, Barroso H, Machado J, Pires I, Cabrita J, Gomes E. Antimicrobial activity of *Terminalia macroptera* root. *J Ethnopharmacol.* 1997; 57: 203-207.
 28. Brandao M G L, Krettli A U, Soares L S R, Nery C G C, Marinuzzi H C. Antimalarial activity of extracts and fractions from *Bidens pilosa* and other *Bidens* species (Asteraceae) correlated with the presence of acetylene and flavonoid compounds. *J Ethnopharmacol.* 1997; 57:131-138.
 29. Estevez-Braun A, Estevez-Reyes R, Moujir L M, Ravelo A G, Gonzalez A G. Antibiotic activity and absolute configuration of 8S-heptadeca-2(Z),9(Z)-diene-4,6-diyne-1,8-diol from *Bupleurum salicifolium*. *J Nat Prod.*1994; 57:1178-1182.
 30. Hufford C D, Jia Y, Croom E M, Jr, Muhammed I, Okunade A L, Clark A M, Rogers R D. Antimicrobial compounds from *Petalostemum purpureum*. *J Nat Prod.* 1993; 56:1878-1889.
 31. Habtemariam S, Gray A I, Waterman P G. A new antibacterial sesquiterpene from *Premna oligotricha*. *J Nat Prod.*1993; 56:140-143.
 32. De Pasquale R, Germano M P, Keita A, Sanogo R, Iauk L. Antiulcer activity of *Pteleopsis suberosa*. *J Ethnopharmacol.*1995; 47:55-58.
 33. Ivanovska N, Philipov S, Istatkova R, Georgieva P. Antimicrobial and immunological activity of ethanol extracts and fractions from *Isopyrum thalictroides*. *J Ethnopharmacol.*1996; 54:143-151.
 34. Afolayan A J, Meyer J J M. The antimicrobial activity of 3, 5, 7-trihydroxyflavone isolated from the shoots of *Helichrysum aureonitens*. *J Ethnopharmacol.*1997; 57:177-181.
 35. Miller E G, Porter L J, Binnie H W, Guo Y I, Hasegawa S. Further Studies on the anticancer activity of *Citrus limonoids*. *J. Agric. Food Chem* 2004; 52: 4908-4912.
 36. Faulks M, Southon S. Carotenoids, Metabolism and Disease. In: Wildman R.E.C. (Ed.). Handbook of Nutraceuticals and Functional Foods. CRC Press, Florida, USA 2001.
 37. Gardner P T, White T A C, McPhail D B, Duthie G G. The Relative Contributions of Vitamin C, Carotenoids and Phenolics to the Antioxidant Potential of Fruit Juices. *Food Chemistry* 2000; 68:471-474.
