



## ISOLATION AND IDENTIFICATION OF GRAM -NEGATIVE BACILLI FROM CLINICAL SPECIMEN

Moh. Rizvan<sup>1,2</sup>, Srashty Sharma<sup>1</sup>, Mukesh Sharma<sup>1</sup>, Swati Tewari<sup>2</sup> and Nayeem ahmad

<sup>1</sup>Department of Microbiology, Maharaja Vinayak Global University, Jaipur

<sup>2</sup>Western Diagnostic Lab, Meerut

### ARTICLE INFO

#### Article History:

Received 4<sup>th</sup> September, 2021

Received in revised form 25<sup>th</sup>

October, 2021

Accepted 23<sup>rd</sup> November, 2021

Published online 28<sup>th</sup> December, 2021

#### Key words:

Gram-Negative Bacilli, Blood agar, MacConkey agar, Biochemical tests, nosocomial infection.

### ABSTRACT

**Introduction:** Gram-Negative Bacilli (GNB) includes numerous organisms but the ones which are known to cause nosocomial infections are *Pseudomonas aeruginosa*, *Acinetobacter baumannii*. This study was undertaken to identify various Gram negative bacteria isolated from patients admitted at a hospital, in Meerut, India. **Material and Method:** A total of 3000 clinical specimens were analyzed. These included 1512(50.4%) urine, 28 (0.9%) pus, 260(8.6%) blood, 159 (5.3%) Sputum, 45(1.5%) CSF and 38 (1.2%) body fluid samples. These samples were inoculated on blood agar, and MacConkey agar. The plates were then incubated at 37°C for 18-24 hours. The clinical isolates obtained were identified using the conventional biochemical tests as per the standardized protocols. **Result:** Majority (1366/3000) of the isolates obtained were Gram-negative bacilli clinical accounting for an isolation rate of 45.5%. Among these GNB, *Escherichia coli* was the most common isolate, accounting for 375 (27.4%) of the total, followed by *Klebsiella pneumoniae* 292(21.3%), *Acinetobacter species* 64(12%), *Enterobacter species* 139 (10%), and *Proteus Species* 134(9.8%). **Conclusion:** GNB are emerging as important opportunistic pathogens and are resistant to most commonly used antimicrobials. Therefore early diagnosis and institution of empirical therapy based on local antibiogram of the institute would reduce mortality and improve patient management.

Copyright©2021 Moh. Rizvan et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### INTRODUCTION

Gram negative bacteria (GNB) are the most common causative agent for morbidities and mortalities in patient admitted in hospitals. Gram-negative bacteria are bacteria that do not retain the crystal violet stain used in the Gram staining method for bacterial differentiation.[1] Gram negative bacteria are characterized by their cell envelopes, which is composed of a thin peptidoglycan cell wall sandwiched between an inner cytoplasmic cell membrane and a bacterial outer membrane. Gram-negative bacteria are found in virtually all environments on Earth that support life.

GNB have great clinical importance in hospitals because they put patients admitted in the intensive care units (ICU) at high risk and lead to high morbidity and mortality[2][3]. These bacteria have a great ability to cause disease in humans and can reach almost all systems in the organism, such as the digestive system, nervous system, urinary system, and

Blood stream, causing diseases ranging from diarrheal gastroenteritis to severe meningitis. GNB colonize the intestines, airways, and skin, which favors the spread of these microorganisms to other parts of the human body, especially in immune compromised individuals.

One of the greatest difficulties of health professionals is to treat nosocomial infections of the lower respiratory tract in which the GNB are involved because they are responsible for a good portion of these infections and are non-responsive to antibiotic therapy due to the high resistance rates and the poor penetration of drugs into the lung parenchyma [4]. The aim of this study was to isolate and identify various bacteria isolated from clinical samples obtained from patients admitted in a hospital located in Meerut city.

### MATERIAL AND METHOD

This study was carried out at western Diagnostic laboratory Meerut from June 2016- July 2021. A total 3000 sample were collected from patients admitted in a hospital in Meerut. The samples included Ascitic fluid 14(0.46%), Blood 260 (8.6%), Pus 794 (26 %), CSF 45 (1.5%), Semen 49 (1.6%), Sputum 159 (5.3%), Pleural Fluid 38 (1.2%), and Urine 1512(50.4%). All the samples were collected at the facility and transported to the laboratory. The samples were immediately inoculated into

\*Corresponding author: Swati Tewari  
Western Diagnostic Lab, Meerut

both differential and enriched media (MacConkey agar and blood agar plates) and then incubate at 37°C for 48 hours. Identification of growth obtained was done based on colony characteristics of the organisms such as beta hemolysis on blood agar, non-lactose fermentation and pigment production (greenish yellow and bluish green pigments) on MacConkey agar. The isolates which showed these characteristics then subcultured onto blood agar to obtain a pure culture and further characterization using the standard biochemical tests.

### Gram Staining and Microscopy

Gram staining was performed on colonies from subcultures for the identification of their gram reaction. Specimens were smeared onto clean grease free glass slides and air dried and then heat fixed and Gram stained. The stained slides were examined microscopically under oil immersion lens for bacterial morphology.

### Bacterial Identification

Biochemical tests included oxidase test, citrate utilization test and indole test was performed to test the enzymatic activities of the organisms. Identification was performed with the Vitek 2 compact (BioMérieux Inc. USA) [5] system using GN ID REF21341 (identification-Gram-negative bacteria) cards. All the test procedures were followed according to the manufacturer's instructions.

## RESULT

Total of 3000 sample was collected from suspected patients during the study period. Of these 1536 (51%) samples were from the male patients and 1464 (49%) were from the female patients. Age of the study population ranged from 1 year to 95 years.

Of the 3000 samples inoculated, Gram negative bacilli were isolated from 1366 (45.5%) samples and Gram positive from 280 (9.3%) samples. 1354 (45%) samples were found to be sterile and no growth was obtained on the medium. The percentage of gram negative isolate in various clinical samples is given in table 1.

**Table 1** Number of isolates, clinical specimen wise; n (%)

Specimen	Total gram-negative bacilli	Percentage (%)
Ascitic Fluid	2	0.14
Bile	5	0.36
Bilateral drain L	2	0.14
Bilateral drain R	2	0.14
Blood	42	3
Cornia	3	0.21
Drain	1	0.07
Drain2	1	0.07
ET Tube	6	0.43
FLUID	4	0.29
Folycil catheter	2	0.14
Fungal	1	0.07
Hvs	4	0.43
Pus	286	20.9
Semen	7	0.51
Swab	5	0.36
Sputum	71	5.1
Urine	891	65.2
Pleural Fluids	3	0.21
Centerline Tip	11	0.80
Throat	2	0.14
Tissue	8	0.58
Tracheal aspirate	7	0.51
Total	1366	

Out of total 1366 Gram negative bacilli, majority of the specimen n=375 (27.4%) were identified as *Escherichia coli*, 292(21.3%) were identified as *Klebsiella pneumoniae*, 164 (12%) were identified as *Acinetobacter Speices*, 139 (10%) were identified as *Enterobacter Species*, 134 (9.8%) were *proteus Species*, 132 (9.6%) were *Citrobacter species*, 69 (5%) were *Serratia marcescens* and 61 (4.4%) were identified as *Pseudomonas aeruginosa*.

## DISCUSSION

In most resource limited settings the emergence and spread of gram negative pathogens is one of the major challenges for the provision of good quality health service in hospitals. Successful management of patients with different kinds of infectious diseases depends on the isolation and identification of the bacterial pathogens. It is well articulated that Gram-negatives are predominantly isolated in different clinical specimens. They are an important cause of nosocomial infections (sepsis, pneumonia, and meningitis) and generally cause severe disease [6]

In the present study, the overall proportion of Gram negative bacilli was 45.5%. Which is similar (45.9%) to the study of Shailpreet Sidhu *et al.*; [7], Ashish Bajaj *et al.*; [8]. Other studies have also reported Gram-negative bacteria as the most common cause of BSIs. [9,10]

In this study *Escherichia coli*, (27.4%) was the most predominant isolate followed by *Klebsiella pneumoniae* (21.3%). The results of this study are in line with the previous. [11].

In our study 12% of the isolates were of *Acinetobacter spp*, 9.8% were *Enterobacter Species*, 9.6% were *Proteus Species* and 5% were *Citrobacter species*. Similar results have been reported in previous studies from India [12, 13].

In the present study, majority of the clinical isolates were recovered from, urine, pus, Pleural fluid, Swab, sputum, etc. Specifically, among the urine culture isolates, *E. coli* and *K. pneumoniae* were the major etiologic agents. This finding is in agreement with the other studies [14] [15] [16].

The frequent use of invasive devices in the form of peripheral venous catheter, urinary catheter, central venous catheter and Endotracheal Tube were found in most of the patients. These devices probably could have acted as a source of infection in these patients [17].

## CONCLUSION

It can be concluded from this study that the most commonly isolated GNB are *Escherichia coli*, and *K. pneumoniae*. GNB are important causative agent of the nosocomial infections and are resistant to commonly used antibiotics. More importantly, these organisms have great potential to survive in hospital environment, so effective methods of sterilization and infection control measures should be implemented to reduce mortality and improve patient management.

## References

1. Baron S, Salton MR, Kim KS (1996). "Structure". In Baron S (ed.). Medical Microbiology (4th ed.). University of Texas Medical Branch at Galveston. ISBN 978-0-9631172-1-2. PMID 21413343.

2. Hormozi SF, Vasei N, Aminianfar M, Darvishi M, Saeedi AA.(2018) Antibiotic resistance in patients suffering from nosocomial infections in Besat Hospital. *Eur J Transl Myol.* Jul 10;28(3):7594. [PMC free article] [PubMed]
3. Li XZ, Plésiat P, Nikaido H. (2015)The challenge of efflux-mediated antibiotic resistance in Gram-negative bacteria. *Clin Microbiol Rev.* Apr;28(2):337-418. [PMC free article] [PubMed]
4. Wenzler E, Fraidenburg DR, Scardina T, Danziger LH. (2016)Inhaled Antibiotics for Gram-Negative Respiratory Infections. *Clin Microbiol Rev.* Jul;29(3):581-632. [PMC free article] [PubMed]
5. Vitek 2 compact (bioMérieux Inc. USA)
6. Melkamu Abeb<sup>1</sup>, Senait Tadesse\*, Girum Meseret and Awoke Derbie. (2019)Type of bacterial isolates and antimicrobial resistance profile from different clinical samples at a Referral Hospital, Northwest Ethiopia: five years data analysis,12:568 <https://doi.org/10.1186/s13104-019-4604-6>
7. Shailpreet Sidhu, Usha Arora, Pushpa Devi.(2010) Prevalence of Non fermentative Gram Negative Bacilli In Seriously ill Patients With Bacteraemia .*jk science*, Vol. 12 No. 4, ,168-171
8. Ashish Bajaj<sup>1</sup>, Bibhabati Mishra<sup>2</sup>,(2019) Prevalence of Gram-negative Septicemia in a Tertiary Care Cente,| Volume: 05 | Issue: 01 | Page: 36 - 41. DOI: <https://doi.org/10.46347/jmsh.2019.v05i01.007>
9. Mehta M, Dutta P, Gupta V.(2005) Antimicrobial susceptibility pattern of blood isolates from a teaching hospital in North India. *Jpn J Infect Dis* ;58:174-6.
10. Kaistha N, Mehta M, Singla N, Garg R, Chander J. (2009) Neonatal septicemia isolates and resistance patterns in a tertiary care hospital of North India. *J Infect Dev Ctries* ;4:55-7
11. Moolchandani K., Sastry A.S., Deepashree. (2017)Antimicrobial resistance surveillance among intensive care units of a tertiary care hospital in South India. *J. Clin. Diagn. Res.* ;11:DC01–DC07. doi: 10.7860/JCDR/2017/23717.9247. [PMC free article] [PubMed]
12. KK Lahiri,\* NS Mani, + and SS Purai(2011). *Acinetobacter* spp as Nosocomial Pathogen : Clinical Significance and Antimicrobial Sensitivity, *Med J Armed Forces India.* Jan; 60(1): 7–10Published online 2011 Jul 21. doi: 10.1016/S0377-1237(04)80148-5
13. Nathan A. Ledebouer,a .(2015) Identification of Gram-Negative Bacteria and Genetic Resistance Determinants from Positive Blood Culture Broths by Use of the Verigene Gram-Negative Blood Culture Multiplex Microarray-Based Molecular Assay; *J Clin Microbiol.* Aug; 53(8): 2460–2472.
14. Beyene G, Tsegaye W.(2011) Bacterial uropathogens in urinary tract infection and antibiotic susceptibility pattern in Jimma University specialized hospital, southwest ethiopia. *Ethiop J Health Sci.* ;21(2):141–6
15. Tadesse S, Kahsay T, Adhanom G, (2018). antimicrobial susceptibility profile and predictors of asymptomatic bacteriuria among pregnant women in Adigrat General Hospital, North- ern Ethiopia. *BMC Res Notes.*; 11(1):740.
16. Mukhtar AM, Saeed HA.(2011) Profile of antibiotic sensitivity and resistance of some pathogenic bacteria isolated from clinical specimens in Sudan. *J Sci Technol.*;12:14
17. Dr. Neeraj , Dr. Sarika , Dr. Bella Mahajan.(2016) Isolation and Identification of Non Fermenting Gram Negative Bacilli in A Tertiary Care Hospital Dr. Ruchita Mahajan<sup>1</sup> , Sch. J. App. Med. Sci.; 4(3D):872-876.

**How to cite this article:**

Moh. Rizvan *et al* (2021) 'Isolation And Identification Of Gram -Negative Bacilli From Clinical Specimen', *International Journal of Current Advanced Research*, 10(12), pp. 25616-25618. DOI: <http://dx.doi.org/10.24327/ijcar.2021.25618.5114>

\*\*\*\*\*