



A STUDY ON QUANTITATIVE ESTIMATION OF PROTEINS OF NEW BREEDING LINES OF BOMBYX MORI L

Manjula A.C¹, Prathibha K.Y², Vishwanatha T³, Naganagouda V Kote⁴ and Keshamma E^{5*}

¹Department of Sericulture, Maharani Cluster University, Palace Road, Bengaluru, Karnataka, India

²Department of Botany, Maharani Cluster University, Palace Road, Bengaluru, Karnataka, India

³Department of Microbiology, Maharani Cluster University, Palace Road, Bengaluru, Karnataka, India

^{4,5}Department of Biochemistry, Maharani Cluster University, Palace Road, Bengaluru, Karnataka, India

ARTICLE INFO

Article History:

Received 15th July, 2021

Received in revised form 7th

August, 2021

Accepted 7th September, 2021

Published online 28th October, 2021

Key words:

Bombyx mori L, Bivoltine, Kalimpong-A (KA), Pure Mysore (PM), Proteins

ABSTRACT

Background: Knowledge on biochemical studies in silkworm is very meagre indicating the type and amount of work to be done in future. At the same time, it is interesting to note that different silkworm races reared in laboratory offer an important testing ground for the application of biochemical methods to taxonomic problems

Objective: The present study was designed to conduct with the main purpose to quantitative estimation of proteins in new breeding lines and races viz. Kalimpong-A (KA), B₁₈, Pure Mysore (PM), evolved R₁ and R₂ of *Bombyx mori* L.

Methods: The modified method of Lowry was used for the quantitative estimation of proteins during the different developmental stages of KA, NB₁₈, PM, evolved R₁ & R₂ races, and the protein concentration was expressed as µg/ml to weight of the tissue.

Results: Results revealed that The soluble proteins in different developmental stages of KA, NB₁₈, PM, R₁ and R₂ have revealed comparatively a low protein content in eggs which however reaches a peak in female pupae. Then declines slowly. The soluble protein content in eggs is high in 24h embryos, later decreases and still decreases in 216 h embryos and in instar larvae suddenly increases and reaches a peak in female pupae of 288 h. Highest quantity of protein was noticed in KA, and is followed by NB₁₈, R₁ and R₂ and PM.

Conclusion: The quantitative analysis of proteins revealed low protein content in eggs, reaches peak in female pupae. The bivoltine parents revealed high protein followed by the hybrids and the Pure Mysore.

Copyright©2021 Manjula A.C et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Any change in the protein/enzyme pattern during the development can be considered as being directly determined by gene function. This reflects on alteration in the metabolism of the developing organism. [1] Analyses of different gene enzyme system have revealed differential regulation of gene activation during metabolic changes. Isozymes can be considered to be ideal gene products for investigating the patterns of gene expression during development. They offer a potential tool and a biochemical index to assess the genetic variability in natural populations. [2-5] These studies have quite often been used to follow the developmental variations in

various enzymatic patterns as well as in species comparison. [6] At the same time, analysis made on isozyme differentiation are systems during ontogeny of wide biological value. [7] The isozymes present during different phases of development can be studied in two ways. One is to understand the differential gene action during development and the other is to compare the manifestation of homologous enzymes during developmental stages.

Though the subject of silkworm genetics, cytogenetics and breeding has gained its importance in Rand D programmes of several laboratories, the literature available with reference to isozyme studies is not sufficient to warrant any plausible line of explanation regarding recognition of different races and the process of speciation in the evolution of silkworm fauna. Furthermore, Considerable data is available at present on

*Corresponding author: Keshamma E

Department of Biochemistry, Maharani Cluster University, Palace Road, Bengaluru, Karnataka, India

aspects of genetics, cytogenetics, breeding, physiology, pathology and embryology of silkworm *Bombyx mori*.^[8] However, our knowledge on biochemical studies in silkworm is very meagre indicating the type and amount of work to be done in future. At the same time, it is interesting to note that different silkworm races reared in laboratory offer an important testing ground for the application of biochemical methods to taxonomic problems. With this viewpoints, the present study was undertaken with the main aim to estimate quantitatively proteins in new breeding lines and races of *Bombyx mori L*.

MATERIALS AND METHODS

Silkworm Varieties and Rearing

The pure races of bivoltine Kalimpong-A (KA) spinning oval white cocoons, New Bivoltine-18 (NB₁₈) spinning dumbbell white cocoons and multivoltine Pure Mysore (PM) spinning pointed yellow cocoons of mulberry silkworm *Bombyx mori L*. were selected for the present breeding programme. These races were obtained from their respective seed areas and are reared in cytogenetics laboratory, Jnana Bharathi, Bangalore University.

The disease free layings were prepared as described by^[9], and were incubated at 25°C and relative humidity of 60-70%. On 8th day composite layings were prepared (10-20 layings were prepared 100-200 eggs were collected from each laying). The hatched worms were reared according to the method described by^[10] MS variety of mulberry leaves were used in rearing. The worms were reared in mass upto III instar, after III moult 300 worms were collected in three replicates in order to evaluate the rearing performance. Standard temperature and humidity were maintained in the rearing house.

Breeding

Single and three way crosses were made by using the above said three races. The first single cross involved KA females and PM males. The second single cross involved NB₁₈ females and PM males. During the course of breeding selection was made at the egg, larva, pupa and cocoon stages to fix the desirable traits. F₅ progenies of the respective crosses were back crossed to their respective bivoltine males to improve commercial characters.

Evolutions of New Lines R₁ and R₂

Females of KA and NB₁₈ were crossed with males of PM. The composite layings of F₁ hybrid were brushed and reared under standard laboratory conditions. The selection parameters explained earlier were applied to choose the seed cocoons for the preparation of F₂ layings. The replicates showing higher pupation rate were selected for intra family selection of cocoons. Further, segregation with respect to cocoon colour and built was noticed. Only white oval in case of KA x PM and dumbbell white in case of NB₁₈ x PM qualifying the parameter of selection were chosen for breeding in subsequent generations. The females of F₅ were backcrossed to the males of KA and NB₁₈ respectively in both the lines and reared up to 11 generations. At the end of the 11th generation the lines R₁ and R₂ were extracted with higher ERR than their respective better parents, with shorter larval period and with moderate cocoon productivity character in case of R₁ and R₂.

Breeding Plans I and II											
I			II								
KA	O	O	x	PM	Cto	NB18	O	O	x	PM	Cfo
	+	i-					+	+			
				F1						F1	
				F2						F2	
				F3						F3	
				F4						F4	
F5	x	KA	O	er'		F5x	NB18	Cta	+		
				F1						F1	
				F2						F2	
				F3						F3	
				F4						F4	
				F5						F5	
				F6 (R1)						F6 (R2)	

Quantitative Analysis of Proteins

The modified method of Lowry et al was used for the quantitative estimation of proteins during the different developmental stages of KA, NB₁₈, PM, evolved R₁ and R₂ races. The protein was expressed as µg/ml to weight of the tissue.^[11]

RESULTS AND DISCUSSION

The soluble protein concentration in any developing organism has been shown to depict marked changes. The present observations confirm the same. The soluble proteins in different developmental stages of KA, NB₁₈, PM, R₁ and R₂ have revealed comparatively a low protein content in eggs which however reaches a peak in female pupae. Then declines slowly. The soluble protein content in eggs is high in 24h embryos, later decreases and still decreases in 216h embryos and in I instar larvae suddenly increases and reaches a peak in female pupae of 288 h. Highest quantity of protein is noticed in KA, and is followed by NB₁₈, R₁ and R₂ and PM (Figs. 1-4).

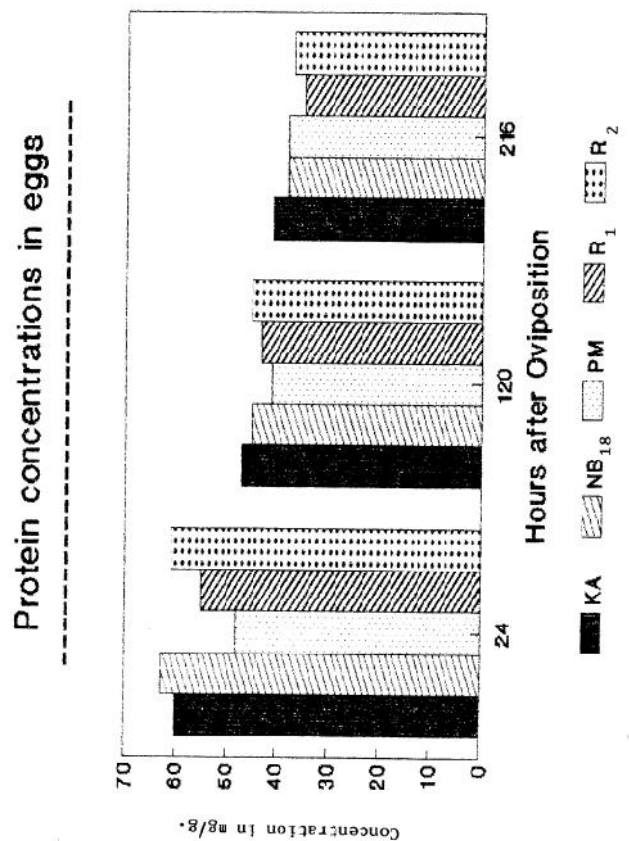


Figure 1

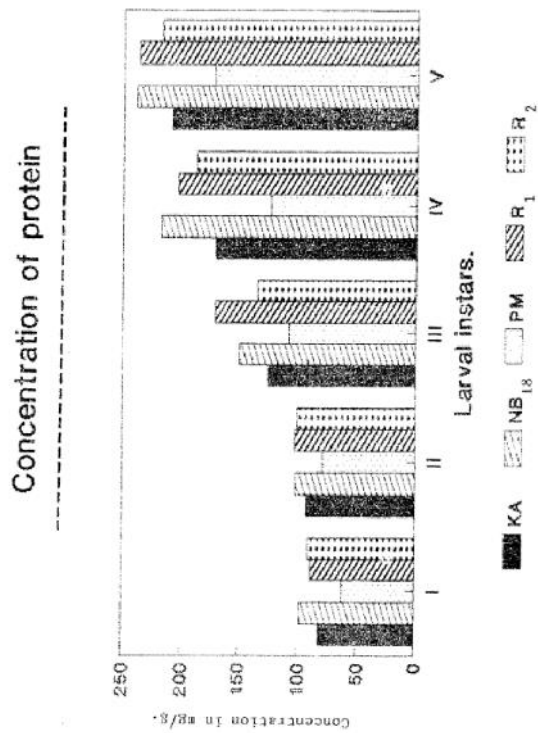


Figure 2

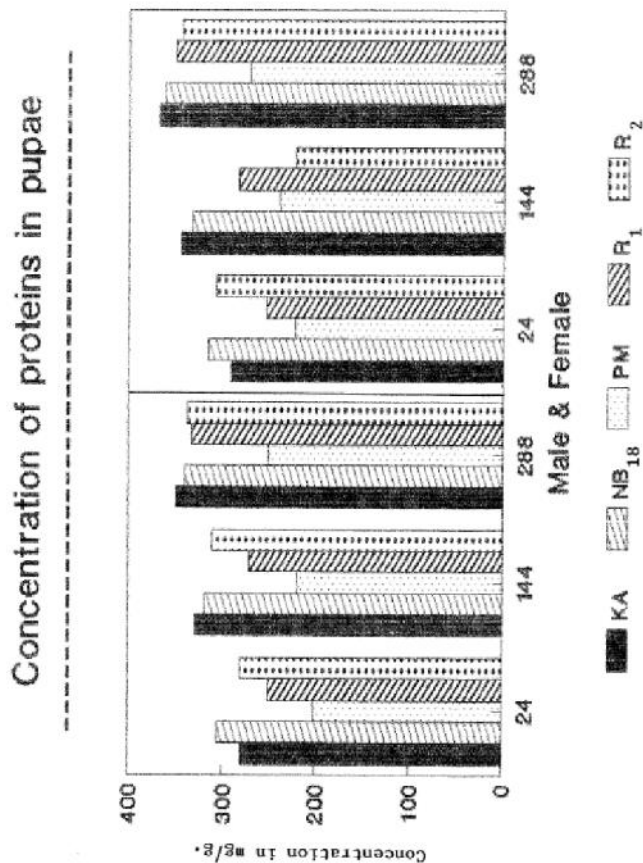


Figure 3

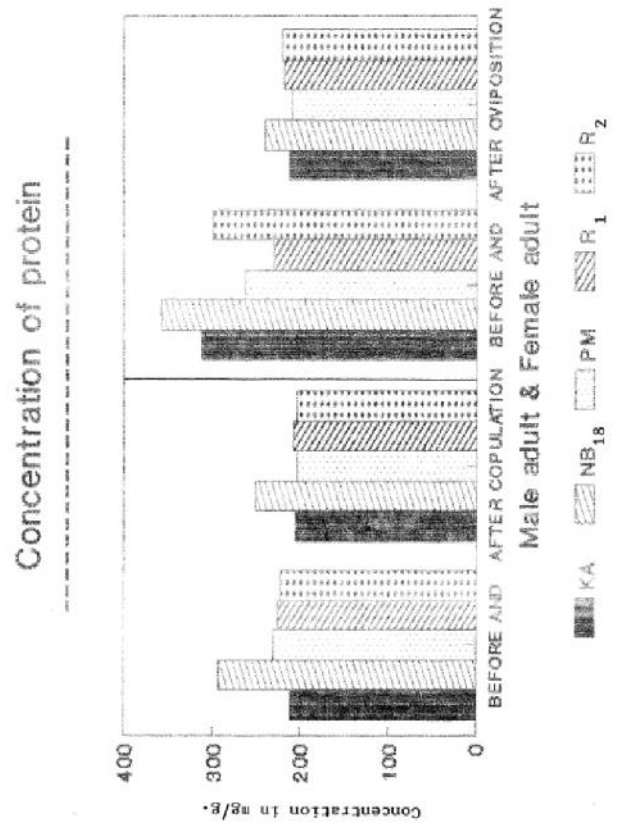


Figure 4

The results of quantitative estimation of protein of new breeding lines and races of *Bombyx mori* L viz. KA, NB₁₈, PM, R₁ and R₂ are presented in figures 1-4. The results revealed that that protein contents in *Bombyx mori* vary according to the race, sex and developmental stage. This is in agreement with similar research involving other species of insects. [12-14] As presented in Figs. 1-4 they are relatively gradual, yet dramatic quantitative changes during the larval and pupal stages and reached lower but relatively constant level of activity in the adult insects. The protein activity in the eggs is high in 24 hours of development whereas it has slightly decreased in 120 and 216 hours. This may be due to the degradation of yolk proteins. Embryogenesis which occurs depend on the degradation of the yolk proteins. [15-19] In larval instars it increases and is shown to be high in pupae which is due to the formation of protein granules in the fat body of an insect. [20]

Many of the quantitative changes that we observe in developing *Bombyx* follow the same trend as reported by some other worker. [21,22] That is, the concentration rises in the growing larvae, reaches peak at some points in the pupal and then taper off to a lower concentration in the adult insect.

CONCLUSION

In conclusion, quantitative analysis of proteins revealed low protein content In eggs, reaches peak in female pupae. The bivoltine parents revealed high protein followed by the hybrids and the Pure Mysore.

References

1. Pasteur, N., & Kastritsis, C. D. (1971). Developmental studies in Drosophila: I. Acid phosphatases, esterases, and other proteins in organs and whole-fly homogenates during development of *D. pseudoobscura*. *Developmental biology*, 26(4), 525-536.
2. Ayala, F. J., Tracey, M. L., Barr, L. G., McDonald, J. F., & Pérez-Salas, S. (1974a). Genetic variation in natural populations of five *Drosophila* species and the hypothesis of the selective neutrality of protein polymorphisms. *Genetics*, 77(2), 343-384.
3. Ayala, F. J., Tracey, M. L., Barr, L. G., & Ehrenfeld, J. G. (1974b). Genetic and reproductive differentiation of the subspecies, *Drosophila equinoxialis* caribbensis. *Evolution*, 24-41.
4. McKenzie, J. A., & Parsons, P. A. (1974). Microdifferentiation in a natural population of *Drosophila melanogaster* to alcohol in the environment. *Genetics*, 77(2), 385-394.
5. Narang, S. (1980). Genetic variability in natural populations, evidence in support of the selectionist view. *Experientia*, 36(1), 50-51.
6. Dickinson, V. J., & Sullivan, D. T. (1975). Gene enzyme systems in *Drosophila*. Results and problems in cell differentiation. Springer-Verlag. Berlin. Heidelberg, New York, 6.
7. Moog, F. Enzyme development, Vol. I (R. Weber Edn.) Acad. Press, N.Y; pp307.
8. Tazima Report on sericulture industry in India (1978). Published by CSB, India. Last accessed on. 2021.
9. Narasimhanna, M. N. (1988). Manual on silkworm egg production.
10. Krishnaswamy, S. (1978). Improved techniques of bivoltine rearing, CSB Publication.
11. Lowry, O. H., Rosebrough, N. J., Farr, A. L., Randall, R. J. (1951). Protein measurement with the Folin phenol reagent. *Journal of biological chemistry*, 193:265-75.
12. Ninmaker, R. A., Mckinnon. (1989). Analysis of proteins variipennins (Diptera Biochem. Physiol., 92B: C.N. Electrophoretic and enzymes in Culicoides: Ceratopogonidae). Comp. 9-16.
13. Liu, T. P., & Dixon, S. E. (1965). Studies in the mode of action of royal jelly in Honey bee development: VI. Haemolymph protein changes during caste Development. *Canadian Journal of Zoology*, 43(5), 873-879.
14. Lensky, Y. (1971). Haemolymph proteins of the honey bee. II. Differentiation during the development of bee workers. *Compar Biochem Physiol*.
15. Engelmann, F. (1979). Insect vitellogenin: identification, biosynthesis, and role in vitellogenesis. In *Advances in Insect Physiology* (Vol. 14, pp. 49-108). Academic Press.
16. Hagedorn, H. H., & Kunkel, J. G. (1979). Vitellogenin and vitellin in insects. *Annual review of entomology*, 24(1), 475-505.
17. Kunkel, J. G., & Nordin, J. H. (1985). Yolk proteins. *Comprehensive insect physiology, biochemistry and pharmacology*, 1, 83-111.
18. Premkumar, D. R. D., Jane, E. P., & Mathavan, S. (1991). Biochemical changes during embryonic development in the aquatic hemipteran bug *Laccotrephes griseus*. *Insect biochemistry*, 21(4), 381-388.
19. Yamamoto, Y., & Takahashi, S. Y. (1993). Cysteine proteinase from *Bombyx* eggs: role in programmed degradation of yolk proteins during embryogenesis. *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry*, 106(1), 35-45.
20. Locke, M. T., & Collins, J. V. (1965). Protein uptake into multivesicular bodies and storage granules in the fat body of an insect. *The Journal of cell biology*, 36(3), 453-483.
21. Chefurka, W., Cambridge, M. A. (1953). Biochemical studies of the blood of gaint silkworm *Hylophora cecropia*, Thesis. Harvard University.
22. Wigglesworth, W. B. (1959). Metamorphosis, polymorphism, differentiation. *Sci. Amer*; 200:100.

How to cite this article:

Manjula A.C *et al* (2021) 'A Study On Quantitative Estimation Of Proteins of New Breeding Lines Of Bombyx Mori L', *International Journal of Current Advanced Research*, 10(10), pp. 25380-25383. DOI: <http://dx.doi.org/10.24327/ijcar.2021.25383.5068>
